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4-Quinazolinones: synthesis and reduction of prostaglandin E₂ production

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Abstract

We synthesized and evaluated the anti-inflammatory activity of a series of 4-quinazolinone derivatives. Two approaches were used to yield the title compounds. A first group of quinazolinone derivatives was obtained by the appropriate substituted anthranilates. A second group of quinazolinone compounds was prepared through the benzoxazin-4-ones intermediate. The pharmacological results reveal that the synthesized derivatives exhibit a significant anti-inflammatory effect in an experimental ocular inflammation model. In fact, all the tested compounds lowered the prostaglandin E₂ (PGE₂) production with respect to the control group ($P < 0.05$). The 3-cyclohexyl-6-chloro-quinazolin-4(3H)-one and 3-cyclohexyl-quinazolin-4(3H)-one derivatives were the most active compounds. These compounds significantly reduced PGE₂ levels even more than the reference drug tolmetin and significantly lower protein concentration and polymorphonuclear leukocytes number compared to the control group ($P < 0.05$). Therefore, these compounds may be useful to prevent ocular inflammatory reactions. © 1999 Elsevier Science S.A. All rights reserved.

Keywords: Quinazolinones; Synthesis; Antiinflammatory; Prostaglandins; Ocular model

1. Introduction

A large number of compounds with a 4-quinazolinonic structure have been synthesized for biological screening, and a variety of interesting pharmacological properties were observed [1]. The most important activities regard their anticonvulsant [2–5] and their anti-inflammatory actions [6–10]. Moreover, several 4-quinazolinone compounds have shown analgesic [11] and antihypertensive [12,13] activities. Recently, a centrally 5-HT_{2A} antagonist activity [14] and a good affinity for cholecystokinin CCK-B receptors [15,16] have been shown from different derivatives with quinazolinonic structure.

As part of an ongoing research project regarding the development of substances with anti-inflammatory activity [17] we have synthesized a series of 4-quinazolinone derivatives with different substituents at the 2, 3 and 6 positions in order to evaluate the decrease of PGE₂ levels after treatment with arachidonic acid (AA).

Interaction of AA with cyclooxygenase produces initially the cyclic endoperoxide prostaglandin G₂ (PGG₂) and thence, through its peroxidase activity, to prostaglandin H₂ (PGH₂). The action of prostaglandin endoperoxide isomerase on PGH₂ produces PGE₂, an important mediator on the inflammatory process [18].

2. Results and discussion

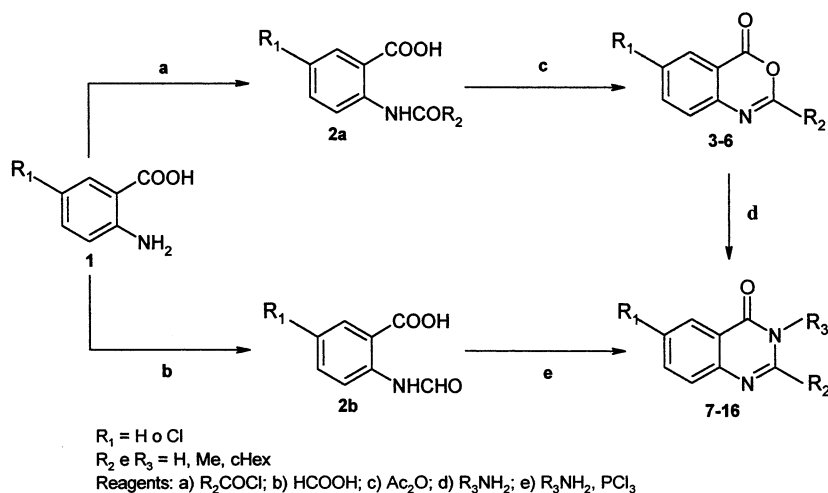
2.1. Chemistry

The synthetic route to the quinazolin-4(3H)-one derivatives is summarized in Scheme 1.

The anthranilic acids (**1**) were coupled with the appropriate acid chloride generating the corresponding substituted anthranilates (**2a**), which underwent cyclization by treatment with boiling acetic anhydride to form the intermediate benzoxazin-4-ones (**3–6**). The substituted benzoxazinones by treatment with ammonia or methylamine solutions yielded the quinazolinone derivatives **7–12**. To prepare the compounds **13–16** we followed a different synthesis method. The two intermediate anthranilates (**2b**), obtained by refluxing the acids

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Scheme 1. Synthetic routes to benzoxazinones (3–6) and quinazolin-4(3H)-ones (7–16).

(1) in benzene with 98% formic acid, were treated with methylamine or cyclohexylamine and subsequently with phosphorus trichloride, to yield the final derivatives **13–16**. The structures of the synthesized compounds were confirmed by elemental analysis, IR and ¹H NMR. Structures, yields, melting points and recrystallization solvents of the benzoxazinones and the quinazolinones are presented in Tables 1 and 2, respectively.

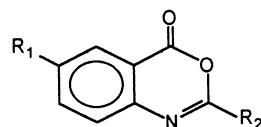
2.2. Pharmacology

The anti-inflammatory activity of the synthesized 4-quinazoline derivatives was evaluated in an ocular inflammation model previously described [19]. An inspection of pharmacological data, reported in Table 3, reveals that all the compounds tested lowered the PGE₂ production with respect to the control group ($P < 0.05$).

We also evaluated the activity of benzoxazin-4-ones intermediate compounds but we did not record any activity. This result may be explained by the fact that

benzoxazin-4-ones in aqueous environment undergo a rapid hydrolytic reaction of the lactonic ring, as demonstrated by high performance liquid chromatographic analysis (data not shown). The 3-cyclohexyl-6-chloroquinazolin-4(3H)-one (**16**) and 3-cyclohexylquinazolin-4(3H)-one (**14**) derivatives were the most active compounds. In fact, these compounds significantly reduced PGE₂ levels even more than the reference drug with 5.5- and 3.8-fold decrease, respectively. In addition these compounds showed a significant reduction in protein concentration, and PMNS number compared to the control group ($P < 0.05$). The compounds **10** and **12** also reduced the PGE₂ production even more than the reference drug with a 3.3- and 2.5-fold decrease, respectively. However, these compounds showed a non-significant reduction in protein concentration, and PMNS number with respect to the control group ($P > 0.05$). Regarding the compounds **7**, **8**, **9**, **11**, **13**, **14** and **15** we observed a decrease of PGE₂ production following arachidonate administration with respect to the control

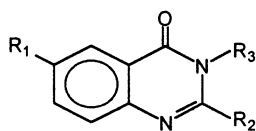
Table 1
Structures, yields and melting points of the prepared benzoxazinone derivatives ^a



Compound	R ₁	R ₂	Yield (%)	M.p. (°C)	Formula
3	H	Me	88	86–87	C ₉ H ₇ NO ₂
4	H	cHex	78	67–69	C ₁₄ H ₁₅ NO ₂
5	Cl	Me	92	124–125	C ₉ H ₆ NO ₂ Cl
6	Cl	cHex	85	104–06	C ₁₄ H ₁₄ NO ₂ Cl

^a All compounds were recrystallized from petroleum ether (b.p. 80–100°C).

Table 2
Structures, yields and melting points of the prepared quinazolines derivatives



Compound	R ₁	R ₂	R ₃	Yield (%)	M.p. (°C)	Formula
7 ^a	H	Me	H	91	238–239	C ₉ H ₈ N ₂ O
8 ^a	H	cHex	H	85	225–226	C ₁₄ H ₁₆ N ₂ O
9 ^a	Cl	Me	H	90	282–283	C ₉ H ₇ N ₂ OCl
10 ^a	Cl	cHex	H	87	262	C ₁₄ H ₁₅ N ₂ OCl
11 ^a	H	cHex	Me	88	111–112	C ₁₅ H ₁₈ N ₂ O
12 ^a	Cl	cHex	Me	80	155–156	C ₁₅ H ₁₇ N ₂ OCl
13 ^b	H	H	Me	56	106	C ₉ H ₈ N ₂ O
14 ^a	H	H	cHex	48	114	C ₁₄ H ₁₆ N ₂ O
15 ^b	Cl	H	Me	52	148	C ₉ H ₇ N ₂ OCl
16 ^a	Cl	H	cHex	41	158	C ₁₄ H ₁₅ N ₂ OCl

^a Recrystallized from ethanol 95°C.

^b Recrystallized from cyclohexane.

group. These compounds were less active than the reference drug tolmetin. In addition these derivatives were not able to reduce the protein concentration and PMNS number compared to the control group.

It has been well demonstrated [19] that topical administration of arachidonic acid causes a production of PGE₂ and leukotriene B₄ (LTB₄) in rabbit aqueous humor as well as a breakdown of the blood–aqueous barrier. LTB₄ is a potent chemotactic agent for PMNs. The treatment with some of the synthesized compounds decreased the number of PMNs in the aqueous humor but not in significant measure, except for compounds **14** and **16**. This could be due to the presence of LTB₄ whose biosynthesis could be inhibited in a different way from the drugs tested. Furthermore, the presence of proteins in the aqueous humor indicates a breakdown of the blood–aqueous barrier which was significantly contrasted by compounds **16** and **14**.

The pharmacological results showed that the presence of the cyclohexylic group was important for the anti-inflammatory activity with respect to the methyl group. In particular, the activity is greater when cycloaliphatic substituent was found in position 3 rather than in position 2. The improvement in the activity of the 3-cyclohexyl-derivatives was not due to the blocking of the tautomerism of the –NH–CO– group as demonstrated by the lower activity of the corresponding 3-methyl-derivatives **12**, **11**, **5** and **13**. In all synthesized derivatives the introduction of a

chlorine atom in position 6 caused a remarkable increment of the activity with respect to the non-substituted analogues. The decrease of PGE₂ levels after drug treatment is particularly interesting because this mediator plays an important role in the ocular inflammation process. The results of this study suggest that the compounds 3-cyclohexyl-6-chloro-quinazolin-4(3H)-one (**16**) and 3-cyclohexyl-quinazolin-4(3H)-one (**14**) may be employed topically to prevent ocular inflammatory reactions.

Table 3

Prostaglandin E₂ levels, protein concentration and number of polymorphonuclear leukocytes in the aqueous humor of albino rabbit 2 h after arachidonate instillation in the eye pre- and post-treated with quinazolin-4(3H)-ones (**7–16**) or reference drug tolmetin

Compound	PGE ₂ ^a (ng/g)	PMNs ^a (PMNs/mm ³)	Protein ^a (mg/ml)
7	7.60 ± 1.50 *	1480 ± 220	24.5 ± 5.1
8	4.00 ± 0.96 *	1420 ± 300	24.6 ± 2.9
9	6.50 ± 1.34 *	1450 ± 195	24.8 ± 3.1
10	0.75 ± 0.02 *	1350 ± 200	23.2 ± 2.7
11	4.15 ± 0.95 *	1380 ± 420	24.1 ± 4.1
12	1.00 ± 0.15 *	1390 ± 320	23.6 ± 2.8
13	6.00 ± 1.35 *	1400 ± 250	24.2 ± 3.1
14	0.65 ± 0.15 *	1200 ± 250 *	20.3 ± 3.1 *
15	5.50 ± 0.91 *	1380 ± 400	24.5 ± 6.1
16	0.45 ± 0.11 *	1100 ± 200 *	19.5 ± 2.8 *
Tolmetin	2.50 ± 0.58 *	700 ± 150	10.1 ± 4.1
Control	22.2 ± 4.31	1500 ± 200	24.7 ± 3.7

^a n = 8 eyes.

* P < 0.05 with respect to control group.

3. Experimental

3.1. Chemistry

2-Amino-benzoic acid and 2-amino-5-chloro-benzoic acid are commercially available from Fluka Chimica (Milan). All other chemicals or solvents were of reagent grade.

IR spectra were recorded on a Perkin–Elmer 1720 spectrophotometer as KBr disks. Melting points were determined with a Büchi 510 apparatus and are not corrected. NMR spectra were performed on a Varian Inova 200 MHz spectrometer. Elemental analyses for C, H and N were obtained on a Carlo Erba 1106 analyser and were within $\pm 0.4\%$ of theoretical values. Thin-layer chromatography (TLC) was carried out on Merck Silica Gel 60 F₂₅₄ (0.25 mm thickness). Column chromatography was performed by the flash procedure.

3.1.1. General procedure for the synthesis of benzoxazinones (3–6)

The intermediate benzoxazin-4-ones **3–6** were prepared according to the literature method [20]. A mixture of the opportune anthranilate derivative of general structure **2a** (50 mmol) and acetic anhydride (45 ml) was refluxed for 3 h. The solution was concentrated under reduced pressure until an oily residue was obtained, which, after cooling at room temperature (r.t.), solidified. IR (cm⁻¹): 1735–1760 (C=O), 1590–1600 (C=N) (see Table 1).

3.1.2. General procedure for the synthesis of quinazolin-4(3H)-ones (7–12)

The title compounds were prepared according to a literature method [21]. A mixture of opportune benzoxazinone derivative **3–6** (40 mmol) in absolute ethanol (20 ml) was treated with an excess of NH₃ 33% solution (40 ml) or methylamine 40% solution (40 ml) and left at r.t. for 48 h. The solid, obtained as light-thin crystals, was filtered, dried, and crystallized by opportune solvent and purified by flash chromatography (toluene:methanol, 95:5 v/v). IR (cm⁻¹): 1665–1700 (C=O), 1590–1600 (C=N) (see Table 2).

3.1.2.1. 2-Methyl-4(3H)-quinazolinone (7). ¹H NMR (DMSO-*d*₆): δ 2.36 (s, 3H, CH₃), 7.41–8.10 (m, 4H, ArH), 12.22 (s, 1H, NH).

3.1.2.2. 2-Cyclohexyl-4(3H)-quinazolinone (8). ¹H NMR (CDCl₃): δ 1.40–2.09 (m, 10H, CH₂-cHex), 2.69–2.80 (m, 1H, CH-cHex), 7.42–8.31 (m, 4H, ArH), 11.78 (s, 1H, NH).

3.1.2.3. 6-Chloro-2-methyl-4(3H)-quinazolinone (9). ¹H NMR (DMSO-*d*₆): δ 2.35 (s, 3H, CH₃), 7.57–8.00 (m, 3H, ArH), 12.38 (s, 1H, NH).

3.1.2.4. 6-Chloro-2-cyclohexyl-4(3H)-quinazolinone (10). ¹H NMR (DMSO-*d*₆): δ 1.17–1.93 (m, 10H, CH₂-cHex), 2.51–2.56 (m, 1H, CH-cHex), 7.59–8.01 (m, 3H, ArH), 12.24 (s, 1H, NH).

3.1.2.5. 2-Cyclohexyl-3-methyl-4(3H)-quinazolinone (11). ¹H NMR (CDCl₃): δ 1.59–2.13 (m, 10H, CH₂-cHex), 2.54–2.75 (m, 1H, CH-cHex), 3.52 (s, 3H, CH₃), 7.11–8.01 (m, 4H, ArH).

3.1.2.6. 6-Chloro-2-cyclohexyl-3-methyl-4(3H)-quinazolinone (12). ¹H NMR (CDCl₃): δ 1.34–1.98 (m, 10H, CH₂-cHex), 2.49–2.70 (m, 1H, CH-cHex), 3.56 (s, 3H, CH₃), 7.41–8.09 (m, 3H, ArH).

3.1.3. General procedure for the synthesis of quinazolin-4(3H)-ones (13–16)

A solution of compound type **2b** (0.1 mol) in pyridine (50 ml) was treated with an excess of methylamine 40% solution (20 ml) or cyclohexylamine (0.1 mol) and PCl₃ (0.12 mol). The mixture was kept in a water bath at 40°C for 90 min. The pyridine excess was then eliminated at reduced pressure, the residue was dissolved by hot HCl (10%). After filtration and cooling at r.t., the filtrate was neutralized by NaOH (20%) and then left to precipitate the final compounds, which were dried and crystallized by opportune solvents. IR (cm⁻¹): 1670–1700 (C=O), 1600–1610 (C=N) (see Table 2).

3.1.3.1. 3-Methyl-4(3H)-quinazolinone (13). ¹H NMR (CDCl₃): δ 3.59 (s, 3H, CH₃), 7.25–8.10 (m, 4H, ArH), 7.39 (s, 1H, CH).

3.1.3.2. 3-Cyclohexyl-4(3H)-quinazolinone (14). ¹H NMR (CDCl₃): δ 1.20–2.03 (m, 10H, CH₂-cHex), 4.74–4.87 (m, 1H, CH-cHex), 7.44–8.33 (m, 4H, ArH), 8.12 (s, 1H, CH).

3.1.3.3. 6-Chloro-3-methyl-4(3H)-quinazolinone (15). ¹H NMR (CDCl₃): δ 3.90 (s, 3H, CH₃), 7.40 (9s, 1H, CH), 7.48–8.11 (m, 3H, ArH).

3.1.3.4. 6-Chloro-3-cyclohexyl-4(3H)-quinazolinone (16). ¹H NMR (CDCl₃): δ 1.01–1.87 (m, 10H, CH₂-cHex), 4.56–4.63 (m, 1H, CH-cHex), 7.61–8.03 (m, 3H, ArH), 8.10 (s, 1H, CH).

3.2. Pharmacology

Arachidonic acid and Tolmetin (used as control drug) were obtained from Sigma Chemical Company (St. Louis, MO, USA). Ketamine HCl (Ketalar[®]) was obtained from Parke-Davis (Milan, Italy). The prostaglandin E₂ (PGE₂) enzyme immunoassay kit was purchased from Amersham Life Science (Milan, Italy) and Coomassie solution was obtained from Pierce (Rockford, IL, USA).

3.2.1. Drug formulations

All drugs tested were formulated at a concentration of 0.02 M in the same isotonic vehicle with a pH of 7.20 (carboxymethylcellulose 1%, NaCl, phosphate buffer). Tolmetin in this condition was soluble whereas all the 4-quinazolone derivatives tested were present as a suspension.

3.2.2. Animals

Female New Zealand albino rabbits (Charles River, Calco, Italy), 1.8–2.2 kg, free of any signs of ocular inflammation or gross abnormality, were used. Animal procedures conformed to the ARVO (Association for Research in Vision and Ophthalmology) resolution on the use of animals in research.

3.2.3. Ocular inflammation

Ocular inflammation was induced as described previously [19] by topical administration (50 μ l) of 0.5% sodium arachidonate dissolved in phosphate-buffered saline (pH 7.4). Drug formulations (50 μ l) were instilled into the conjunctival sac 180, 120, 90 and 30 min before induction of ocular inflammation by arachidonate (pre-treatment) and then again 60 min thereafter (post-treatment). Two hours after the arachidonate instillation the rabbits were anesthetized by intravenous injection of 20 mg/kg of Ketamine HCl. Aqueous humor was withdrawn by a tuberculin syringe and divided into three aliquots in order to evaluate the content of PGE₂ by enzyme immunoassay, the protein concentration by Coomassie solution and polymorphonuclear leukocyte (PMNs) levels by an improved Neubauer chamber. The experimental plan included a group of four animals for each drug tested including the reference group (treated with the reference drug tolmetin) and the control group that received no treatment.

3.2.4. Statistical analysis

Results are expressed as the mean \pm SD Student's *t*-test was used to evaluate the significance between the groups of animals. The statistical significance was fixed at $P < 0.05$.

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